NANOLIVE
Looking inside life

Short sample preparation guide for Nanolive field demos

USE SUITABLE LABWARE

Nanolive suitable labware is IBIDI µ-dish 35mm or equivalent:
1.5# and 170µm bottom thickness (Ex. Eppendorf, Cellvis, Iwaki, Nunc™, WPI)

µ-Dish 35mm, high
(Polymer Coverslip,
Tissue culture treated)
ref. 81156

µ-Dish 35mm, high
Glass Bottom
ref. 81158

µ-Dish 35mm, low
Polymer Bottom
Tissue culture treated
DIC lid compatible
ref. 80136

ADD 1ML–2ML MEDIUM

Assure that the medium level is flat (no meniscus) – avoid bubbles.

KEEP THE MEDIUM AS CLEAN AS POSSIBLE

Change medium before imaging – assure no floating debris or dead cells.
Use prewarm medium – bring some with you to the demo.

AIM FOR 60% CELL CONFUENCY

Allow strong cell adhesion for adherent cells.

FAQ'S

1. Can I use fixed samples? **Answer:** Yes.
2. Can I use tissues slices? **Answer:** Yes, thin (up to 10µm thickness) usually work.
3. Which fluorophores can I use? **Answer:** almost any, contact your demo organizer for more details.
4. Can I use a dish coating? **Answer:** Yes. Thin coating is suitable. Fibronectin or similar is ideal. Some more dense or less transparent coatings can affect image quality (ex. matrigel).
5. Can my sample be incubated at controlled temperature, CO2 and humidity? **Answer:** Yes.
6. Can I image co-culture? **Answer:** Yes. For adherant cells with non adherent: change medium before adding non adherent cells and start with low confluency of non adherent cells (bring more with you).
7. Two non adherent cells lines: use fluorescent marker to assure distinguishing of the two cell lines.

For further questions contact your demo organizer at **Sanela.vellino@nanolive.ch**.